ACADEMIC Journal of Science

ISSN: 2165-6282

UniversityPublications.net

Home

Copyright

About AJS

Editorial Board

Table of Contents

Author Index

Search

Help

Journal of Science

Academic

CD-ROM ISSN: 2165-6282

2013 Volume 02, Nümber 01

This issue, or parts thereof, may not be reproduced in any form or by any means, electronic or mechanical, including photocopying, recording or any information storage and retrieval system now known or to be invented, without written permission from the Editor(s) or from the UniversityPublications.net.

Copyright © 2012 UniversityPublications.net

29/10/2013 13:12

Search

ACADEMIC Journal of Science

ISSN: 2165-6282 UniversityPublications.net

Home

Copyright

About A15

Editorial Board

Author Index

Search

Help

Editorial Board

Associate Editors

Nancy Mathes, ISSI, USA

Joseph Bonnici, Central Connecticut State University, USA

Joseph Azzopardi, University of Malta, Malta

Mary Allen, UniversityPublications.net, USA

Consulting Editors

Peter Mini, Bryant University, USA

Gene Gulledge, University of Arkansas - Monticello, USA

Bill Hamby, Indiana Wesleyan University, USA

Michael McKinney, Jamestown College, USA

Anthony Lopez, UniversityPublications.net, USA

David Moore, UniversityPublications.net, USA

Conferences Advisory Board

Kathryn LaFever, Miami University, USA

Rose Marie Azzopardi, University of Malta, Malta

Lukas Jirsa, Charles University, Czech Republic

Henry Greene, Central Connecticut State University, USA

Sangeeta Khorana, Aberystwyth University, UK

Khoon Koh, Central Connecticut State University, USA

Debopriyo Roy, University of Aizu, Japan

S. Serhat Serter, Anadolu University, Turkey

Jim Cross, Leeds Trinity & All Saints College, UK

International Editorial Board ¹

Dustin Morrow, Temple University, USA

Christopher J. Greig, University of Windsor, Canada

Carlos Liard-Muriente, Central Connecticut State University, USA

Suzanne Maniss, University of Tennessee at Martin, USA

Anna Gemra, University of Wroclaw, Poland

Josephine Etowa, University of Ottawa, Canada

Vida Davoudi, Lonestar College, USA

Brian Phillips, Grand Valley State University, USA

Kavita Daiya, George Washington University, USA

H. Dean Buttram, III, Jacksonville State University, USA

S. Serhat Serter, Anadolu University, Turkey

Caroline Brandt, The Petroleum Institute, Abu Dhabi, UAE

Danielle Twigg, Griffith University, Australia

A. Helene Robinson, St. John's University, USA

Shannon Melideo, Marymount University, USA

Irina Moreland, University of Colorado at Denver, USA

Tao Zeng, WIlfrid Laurier University, Canada

Igor Pustylnick, Humber College, Canada

Larry Riggs, Butler University, USA

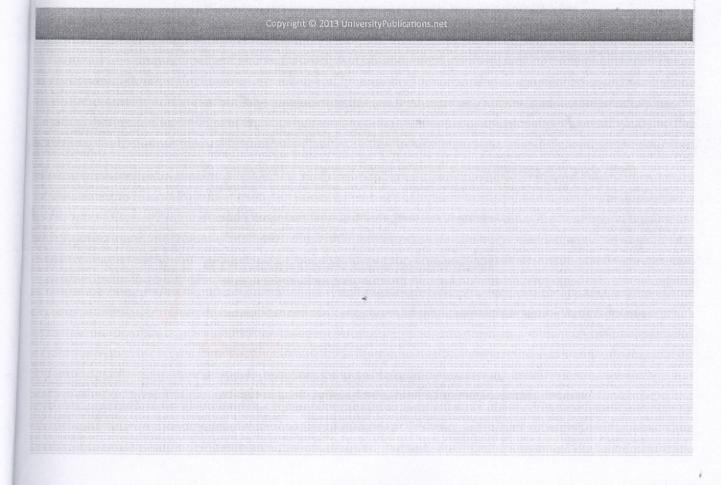
Shehla Burney, Queens University, Canada

Maria Assif, University of Toronto, Canada

Lyndon B. Carew, University of Vermont, USA

María Rosalía Vicente, University of Oviedo, Spain Radka Wildova, Charles University, Czech Republic Gurmak Singh, University of Wolverhampton, UK Monika Raesch, Suffolk University, USA Ephie Konidaris, McGill University, Canada Bulent Tarman, Selcuk University, Turkey Shoji Ohga, Kyushu University, Japan Ardian Greca, Georgia Southern University, USA Hasan Ozdemir, Istanbul University, Turkey Ruxandra Serbanescu, University of Toronto, Canada Reema Khurana, Institute of Management Technology, India Rina Devnita, Padjadjaran University, Indonesia Pattabhi Sitaram, Purdue University Calumet, USA Rich Snow, Embry-Riddle Aeronautical University, USA Michael Quinn, Bentley University, USA Juan C. Barrera, Elmhurst College, USA Kevin E. Dow, University of Alaska Anchorage, USA Marita Naudé, Curtin University, Australia Christopher Kubik, Colby-Sawyer College, USA Sisira R.N.Colombage, Monash University, Australia Daniel E. May, Harper Adams University College, UK Sofia Daskou, Hellenic American University, Greece Chrysost Bangaké, University of Orleans, France Saranyapong Thiangtam, Bangkok University, Thailand Adugna Lemi, University of Massachsetts Boston, USA Aizzat Mohd. Nasurdin, Universiti Sains Malaysia, Malaysia David S. Kung, University of La Verne, USA Rima Aboudan, United Arab Emirates University, UAE

¹This list does not include the ad hoc reviewers. Given the diversity of papers in multidisciplinary publishing, it is common practice for the review process in such journals to rely on ad hoc reviewers whose research interest is in line with the research topic of a submitted manuscript. The expertise of these reviewers is invaluable to the review process.



Search

ACADEMIC Journal of Science

ISSN: 2165-6282 UniversityPublications.net

Copyright

About AJS

Editorial Board

Table of Contents

Author Index

Search

Help

Table of Contents (Volume 02, Number 01) (Download in PDF 🖎)

- Experimental Investigation of the Effect of Drying Air Relative Humidity in Drying Kinetics of Kiwi Soner Şen and Selçuk Darici
- An Experimental Investigation of Effect on Engine Emission of Addition at Dual Fuel Mode of Hydrogen
 H. Köse and M. Ciniviz
- Evaluation of the Cyanogenetic Potential of Selected Plants: I. Legumes

 Juliana Janet R. Martin-Puzon and Maria Socorro M. Mendoza
- Determination of the Effects of Safflower Biodiesel and Its Blends with Eurodiesel on Engine Emissions in a Single Cylinder Diesel Engine
 Abdullah Engin Özçelik, Hüseyin Öğüt, Ergün Çitil and Seda Şahin
- The Usability of Abs/Pc Composit Materials as Gear Material and the Effect of the Heat Accumulated on the Tooth Surface Rifat Yakut and Hayrettin Düzcükoğlu
- Transient Conjugated Heat Transfer in a Circular Pipe Ali Ateş and Şefik Bilir
- Leaching of Chalcopyrite Concentrate (CuFes₂) in Nitric Acid (HNO₃) Solution Tevfik Ağaçayak and Ali Aras
- Determination of Leaching Conditions of Galena Concentrate in Acid Medium with Chloride Ions Ali Aras and Tevfik Ağaçayak
- The Usage of Miscanthus (Giganteus) as Vegetable Fiber in the Production of Reinforced Concrete

 Beams

Hicran Açikel, Necati Yalçin and Durmuş Ali Açike

- Anthropometric Measures of Student Athletes in Selected Malaysian State Sports Schools
 Ong Kuan Boon, Hishamuddin Ahmad, Sanmuganathan Jegathesan and Lee Ai Choo
- Space Vector Pwm Controlled Single Phase Line Interactive Ups Hüseyin Doğan and Ramazan Akkaya
- Effect of Chopped Kevlar to Wear and Braking Performance Mahmut Ünaldi and Recai Kuş
- Coconut Shell Activated Carbon as an Alternative Sedimentation Agent in Water Purification
 System
- Suparno and Fina Mutiara Besty
- Fatty Acid Composition of Muscle Lipids of Five Sheep Breeds

 Ali Karabacak, Saim Boztepe, Gökalp Ozmen. Guler and Abdurrahman Aktumsek



COCONUT SHELL ACTIVATED CARBON AS AN ALTERNATIVE SEDIMENTATION AGENT IN WATER PURIFICATION SYSTEM

Suparno and Fina Mutiara Besty

Yogyakarta State University, Indonesia

An investigation on the use of coconut shell activated carbon as sedimentation agent of dirty water is reported. The aim of the research is to find an alternative substance which can be used as sedimentation agent in a water purification system, which is badly needed in areas hit by natural disaster such as earth quake, flood and landslide. Carbon made of coconut shell was chosen due to its ample availability as a waste product in most places all over Indonesia. This was physically activated by heating it in an oven at 120°C for 30 minutes and chemically activated by soaking it in 40%-80% sulfuric acid, H_2SO_4 , for 24 hours. This coconut shell activated carbon was then mixed with simulated dirty water and the sedimentation velocity of the dirt coagulant was then observed. The results were then compared to those of the well known commercial sedimentation agents for water purification system, aluminum sulfate, $Al_2(SO_4)_3$ and poly aluminum chloride (Al $Cl_3)_n$. The results show that by using coconut shell activated carbon the dirt coagulants are much quicker to sediment compared to those of aluminum sulfate and poly aluminum chloride. By using 1 gram activated carbon in 1 liter dirty water the sedimentation velocity were observed to be ranging from $(3.98 \pm 0.08) \times 10^3$ m/s to $(4.51 \pm 0.03) \times 10^3$ m/s. On the other hand by using aluminum sulfate and poly aluminum chloride the sedimentation velocities of the dirt coagulant are $(2.57 \pm 0.06) \times 10^3$ m/s and $(2.47 \pm 0.06) \times 10^3$ m/s respectively.

Keyword: Coconut shell activated carbon, Sedimentation agent, Coagulant, Aluminum sulfate, Poly aluminum chloride.

Introduction

Geographically Indonesia is located in a ring of fire. Therefore, there are hundreds of tectonic and volcanic earth quakes in a year. Earth quake causes well water to shake and results in dust and dirt on the bottom of the well to disperse throughout the water. Consequently, the well water in the disastrous area is dirty and leaves people without clean water for washing, drinking and cooking. The same problem occurs when an area is hit by flood, tsunami, and landslide. Unfortunately people cannot stay long without clean water. Consuming dirty water may cause the spreading of diarrhea, cholera, and other related diseases, which could endanger people's life. In order to avoid the worst scenario a simple, cheap, and easy way of cleaning dirty water is badly needed.

More than sixty percents of 240 million Indonesian populations live in villages and remote areas. Most of them consume water from the well. Due to bad water management system more than half of people living in urban areas also rely on the well water. When an area is hit by earthquake, flood, tsunami, or landslide the well water becomes extremely dirty. The dirt and dust in the bottom of the well are dispersed in the well water due to the quake, flood, tsunami, and landslide. This dispersion of dirt

belongs to colloidal dispersion which is relatively stable. Ions from minerals in the dirt contribute to electrostatic kinetic stability.[1]-[3] Studies on charged particles have been done extensively using Phase Analysis Light Scattering (PALS).[4]-[8] A review on various techniques for colloidal particle surface charge determination has been published by Suparno.[9] It takes days to weeks to sediment the dirt in the water naturally.

Most people use *aluminum sulfate*, Al₂(SO₄)₃ or *poly aluminum chloride*, (AlCl₃)_n to clean the water.[10] These two substances are not only known as sedimentation or flocculating agent but also known as anti-perspirant and deodorant.[11] They put tens to hundreds grams of *aluminum sulfate* or *poly aluminum chloride* (PAC) in the well and wait for the dirt to sediment to get the clean water. These substances promote coagulation among the dirt and the dirt coagulants are then sediment to the bottom of the well resulting in cleaner water. However, it takes too long to clean dirty water using these two substances. People living in the disastrous areas cannot live for too long with dirty water. Therefore, an alternative substance should be found to accelerate sedimentation of the dirt in the well water.

A research has been done to investigate coconut shell activated carbon as alternative sedimentation agent to clean the water. Coconut shell has been chosen since as a waste product it is available in huge amount all over Indonesia. Most Indonesians consume coconut in their daily life. After the coconut being consumed, the shell was usually thrown away or in some cases being burnt. Massive use of coconut can be found in coconut oil industries. These industries produce a huge amount of coconut shell waste product. It is easy to find piles of coconut shell in and around coconut oil industries and most of them end up being burnt.

The use of coconut shell as activated carbon in various areas [12]-[16] inspired us to make use of it as sedimentation agent. Therefore, an extensive work has been done in an effort to make it a better substance for sedimentation of the dirt inside the water. The coconut shell has to be carbonated in an extremely high temperature in a closed tight carbonation chamber. No air is allowed to flow into the chamber to prevent oxidation that results in ashes. The resulting carbon was then being cooled slowly. The coconut shell carbon was then crust and grind into fine pieces of approximately 50 meshes. These fine pieces of coconut shell carbon were then physically and chemically activated before being used as sedimentation agent. The superiority of coconut shell activated carbon as sedimentation agent was shown by measuring the sedimentation velocity of the dirt coagulants and comparing the results with those of commercial sedimentation agents, aluminum sulfate and poly aluminum chloride.

Carbonation, Carbon Activation, and Sample Preparation

Carbonation process of the coconut shell was done in a closed tight carbonation chamber at 400°C for 2.5 hours. The process of pyrolysis occurs in the chamber resulting in carbon. If the air is flowing into the chamber the oxidation process occurs instead of pyrolysis, resulting in ashes. The resulting coconut shell carbon was then being cooled slowly inside the chamber to avoid contact with air.

The coconut shell carbon was crust and grind to small pieces. These small pieces of carbon were then sifted to 50 meshes. This means that there are 50 pieces of carbon powder standing in a row of one inch. This size is equivalent to 368 micrometer. The sieve to small size is designed to enlarge the surface area and this is useful to accommodate more dirt particles to adsorb onto the surface of activated carbon. Studies on micrometer size colloidal particles may be done using Dynamic Light Scattering.[17]-[19]

The carbon powder was separated into 5 groups and each of them was chemically activated by soaking it in the solution of 40% - 80% sulfuric acid for 24 hours. Sulfuric acid is colorless and soluble in water. It is very strong acid and decomposes protein and lipid. It is commonly used as electrolyte and cleaning agent. The chemical activation using sulfuric acid is designed to destroy organic materials trapped inside the pores of the carbon powder. The trapped materials reduce the capacity of the pores significantly. The destroyed organic materials were then thrown away by rinsing the carbon under flowing clean water. The results were then dried up using oven at 90°C and being kept away from contact with water.

Physical activation was done prior to the research. This was aimed to evaporate water molecules from the carbon pores to ensure maximum capacity of the pores. The maximum capacity of the pores is very essential, since it represents maximum absorption of the dirt in the chamber of pores and maximum adsorption of the dirt on the wall of the pores. The physical activation was done in an oven and for this purpose the temperature of the oven was set to be 120°C for 30 minutes.

Simulated dirty water sample is made by diluting 25 grams of dirt in a liter of clean water. The dirt was made of soil powder. This was stirred for 30 second before being ready to use as sample. A gram of coconut shell activated carbon (chemically activated in 40% of sulfuric acid) was added into a liter of sample and then stirred manually for 30 minutes. The mixture of sample and activated carbon was placed into measuring tube to allow the observation of sedimentation in a relatively long distance.

When the dirt coagulant is falling to the bottom of the measuring tube, at first it experiences accelerated motion due to gravitational force of the earth. In this case the gravitational force, F_g is given by

$$F_g = mg \tag{1}$$

where m is the mass of the falling dirt and g is the acceleration.

However, this motion was challenged by the viscous force that is increasing with the increase of the velocity of the dirt, v. The viscous force for a spherical object is given by

$$F_g = 6\pi \eta R v \tag{2}$$

where η is the viscosity of the solution, R is the radius of the moving object and v is the velocity of the object. The gravitational force causes the dirt to move faster. However the higher the velocity of the falling dirt the higher the viscous force. This means that the two forces will soon reach their equilibrium and at the equilibrium the dirt moves in a constant sedimentation velocity.

The determination of the sedimentation velocity was done by the measurement of sedimentation time, Δt needed by the dirt coagulants to sediment across 10 cm distance, Δx . The average sedimentation velocity, v is calculated by using Equation 3:

$$\overline{v} = \frac{\Delta x}{\Delta t} \tag{3}$$

In order to get more accurate result, the sedimentation time measurements were done 20 times each and the value used in the calculation is the average of 20 measurements. In order to understand the influence of chemical activation to the sedimentation velocity the same process was repeated by using 50% - 80% sulfuric acid activated carbon. Furthermore, to understand the influence of the amount of the carbon to the sedimentation velocity the process was repeated for 1-5 grams of activated carbon mixed with a liter of the sample.

The Performance of Commercial Sedimentation Agents

Commercial sedimentation agents such as aluminum sulfate and poly aluminum chloride are commonly used to clean water in the well. The white crystal aluminum sulfate is more popular compared to poly aluminum chloride. One gram of aluminum sulfate is dissolve in the simulated dirty water sample and stirred for 30 seconds. This was then left for 60 seconds in a measuring tube before the measurement of sedimentation time started. The sedimentation time of the dirt coagulant across 10 cm distance in the sample was measured for 20 times each and the average of these values was used to calculate the sedimentation velocity using Equation 3. This was done repeatedly for 1-5 grams of aluminum sulfate and poly aluminum chloride.

Figure 1 shows the performance of aluminum sulfate and poly aluminum chloride to sediment the dirt in simulated dirty water. By using 1-5 grams aluminum sulfate the sedimentation velocity was

calculated to be in between $(2.57 \pm 0.06) \times 10^{-3}$ m/s to $(2.97 \pm 0.06) \times 10^{-3}$ m/s. These results were relatively higher compared to those of using 1-5 grams poly aluminum chloride which are ranging from $(2.15 \pm 0.05) \times 10^{-3}$ m/s to $(2.47 \pm 0.06) \times 10^{-3}$ m/s. These results quantitatively answer the question why people prefer to use aluminum sulfate than poly aluminum chloride.

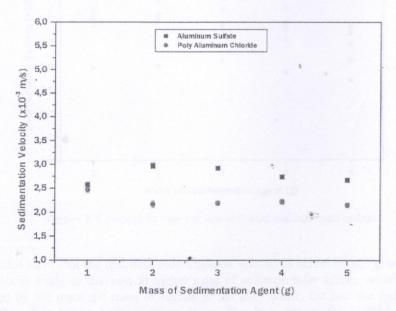


Figure 1. Sedimentation velocity using commercial Sedimentation Agent.

Contribution of Activation to the Increase of Sedimentation Velocity

In order to appreciate the role of chemical activation to increase the sedimentation velocity the data of sedimentation velocity using carbon without being activated and using activated carbon are presented in Figure 2. The mass of carbon for both types is varied from 1-5 grams. As for the activated carbon, there were five groups being identified by the concentration of sulfuric acid, H_2SO_4 used to chemically activate the carbon. The concentrations of sulfuric acid for the five groups of carbon were 40%-80% respectively with 10% increment.

Figure 2 shows that the sedimentation velocity of the dirt coagulants by using carbon without activation is ranging from $(2.68 \pm 0.03) \times 10^{-3}$ m/s to $(3.10 \pm 0.03) \times 10^{-3}$ m/s. On the other hand the best performance was achieved by using 60% sulfuric acid activated carbon. The sedimentation velocity by using 60% sulfuric acid activated carbon is ranging from $(3.80 \pm 0.06) \times 10^{-3}$ m/s to $(4.75 \pm 0.11) \times 10^{-3}$ m/s. There is an increase of approximately 40-50 percents of the sedimentation velocity by using activated carbon. This is understandable since chemical activation destroyed any organic materials blocking the pores and they were flushed during the rinsing process leaving the pores widely open and ready to absorb the dirt from the sample. The more the dirt trapped inside the pores of activated carbon in a short time the quicker the dirt falls to the bottom of the measuring tube. The fewer the dirt trapped inside the pores the slower the dirt falls, since the carbon itself tend to stay at the surface of water due to its specific mass that is less than the specific mass of water. The most likely cause bringing carbon to fall is the dirt coagulant that is trapped inside the pores by the process of absorption and the dirt that is stuck on the surface of carbon by the process of adsorption.

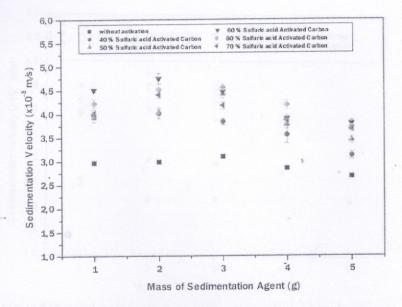


Figure 2. Comparison between non activated and activated carbon.

The data also show that the optimum sedimentation velocity is achieved at 2 grams of sedimentation agent. The velocity tends to decrease at higher mass of sedimentation agents, which is carbon. This is possibly caused by the more the mass of carbon in the dirty water, the less the dirt trapped inside the individual carbon that brings both of them down. The less dirt in the individual carbon the less gravitational force that drags the carbon along with the dirt downs. This means that the equilibrium force condition is achieved at relatively low viscous force, which means relatively low sedimentation velocity.

Comparison of Commercial Sedimentation Agent to Activated Carbon

The performance of various activated carbons as sedimentation agents will not be justified without comparison to the performance of commercial sedimentation agents. This research observed 5 different activated carbons. Originally they are the same carbon that is chemically activated using 5 different sulfuric acid concentrations, which are 40%, 50%, 60%, 70% and 80%. The commercial sedimentation agents used in this research are aluminum sulfate and poly aluminum chloride.

Figure 3 presents sufficient data showing that sedimentation velocities due to various activated carbons are much greater than those of due to commercial sedimentation agents. The difference in sedimentation velocity is significantly high at one gram of sedimentation agents. At this point the highest sedimentation velocity due to commercial sedimentation agent is $(2.57 \pm 0.06) \times 10^3$ m/s which is represented by aluminum sulfate. On the other hand, the highest sedimentation velocity due to in house sedimentation agent, which is activated carbon, is $(3.98 \pm 0.03) \times 10^3$ m/s. This is approximately 55% higher compared to that of commercial aluminum sulfate. Higher sedimentation velocity means quicker to clean the water. This might be caused by sufficiently large space in pores that is capable of absorbing a large amount of dirt and sufficiently large surface area that is capable of facilitating adsorption of the dirt on the surface of the carbon. However another research should be done to uncover the contribution of absorption and adsorption of the dirt individually.

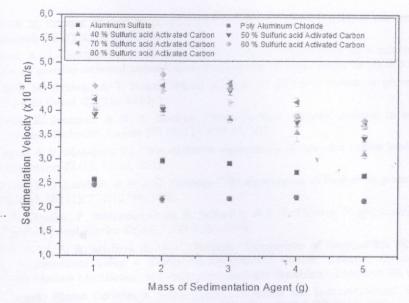


Figure 3. Comparison between commercial sedimentation agent and activated carbon.

Conclusion

From the above discussion it can be concluded that the sulfuric acid activated carbon can be used as an alternative sedimentation agent. In fact it contributes to higher sedimentation velocity compared to those of commercial aluminum sulfate and poly aluminum chloride. Furthermore, it was uncover that chemical activation greatly increases the performance of carbon as a sedimentation agent. A more work should be done to understand the contribution of absorption and adsorption individually to the performance of activated carbon as an alternative sedimentation agent.

Reference

- P. C. Heimens & R. Rajagopalan, Principles of Colloid and Surface Chemistry, 3rd ed., Marcel Dekker, New York, 1997.
- 2. D. F. Evans & H. Wennerstron, The Colloidal Domain Where Physics, Chemistry, Biology, and Technology Meet, Wiley VCH, New York, 1999.
- 3. D. Myers, Surface, Interfaces, and colloids, Wiley-VCH, New York, 1999.
- R. I. Keir, Suparno, J. C. Thomas, "Charging behavior in the Silica/Aerosol OT/Decane System", Langmuir, 18, pp. 1463; 1465, 2002.
- R. I. Keir, A. Quinn, P. Jenkins, J. C. Thomas, J. Ralston, & O. Ivanova, #Electrokinetic Properties of Copper Pthalocyanine Pigment Dispersions\$, Journal of Imaging Science and Technology, 44(6), pp. 528! 533, 2000.
- J. C. Thomas, K. L. Hanton, & B. J. Crosby, #Measurement of the Field Dependent Electrophoretic Mobility of Surface Modified Silica/AOT Suspensions\$, Langmuir, 24(19), pp. 10698! 10701, 2008.
- 7. J. F. Miller, K. Schatzel, & B. Vincent, Journal of Colloid and Interface Science 143, pp. 532! 534, 1991.
- J. F. Miller, B. J. Clifton, P. R. Benneyworth, B. Vincent, I. P. McDonald, & J.F. Marsch, Colloid Surfaces 66, pp. 197! 202, 1992.
- Suparno, "A review on prominent techniques on the determination of colloidal particle surface charge", IJBAS-IJENS, 12(4), pp. 74!77, 2012.
- C. Exley, "Does anti-perspirant use increase the risk of aluminum-related disease, including Alzheimer's disease?", Molecular Madicine Today, 4(3), 107! 109, 1988.

- 11. S. Kvech, & M. Edward, "Solubility controls on aluminum in drinking water at relatively low and high pH", Water Research 36(17), 4356! 4368, 2003.
- A. Aizpuru, A., L. Malhautier, J. C. Roux, & J. J. Fanlo, "Biofiltration of a mixture of volatile organic compounds on granular activated carbon", Biotechnology and Bioengineering 83 (4), 479! 488, 2003.
- M. Michael, M. Brittain, & J. Nagai, "Phase II study of activated carbon to prevent irinotecan-induced diarrhea". J Clin Oncol. 22 (21): 4410! 7.
- M. Eddlesto, E. Juszczak, & N. A. Buckley, "Multiple-dose activated charcoal in acute self-poisoning: a randomised controlled trial". Lancet 371 (9612): 579! 87, 2008.
- 15. R. V. Shende & V. Mahajani, V., "Wet oxidative regeneration of activated carbon loaded with reactive dye". Waste Management 22 (1): 73! 83, 2002.
- G. S. Miguel, S. D. Lambert, & N. J. D. Graham, "The regeneration of field spent granular activated carbons". Water Research 35 (11): 2740! 2748, 2001.
- Supamo, K. Deurloo, P. Stamatelopolous, R. Srivastva, & J. C. Thomas, "Light scattering with single mode fiber collimators", Appl. Optics 33(30), 7200! 7205, 1994.
- I. Takashi, S. Li, A. B. Micheal, & M. C. Richard, "Comparison of Nanoparticle Size and Electrophoretic Mobility Measurements using a Carbon-Nanotube-Based Coulter Counter, Dynamic Lifgt Scattering, Transmission Electron Microscopy, and Phase Analysis Light Scattering", Langmuir 20, 6940! 6945, 2004.
- J. C. "Thomas, Photon Correlation Spectroscopy: Technique and Instrumentation" in "Photon Correlation Spectroscopy: Multicomponent Systems", Schmitz, KS, Proc. SPIE 1430, 2! 18, 1991.